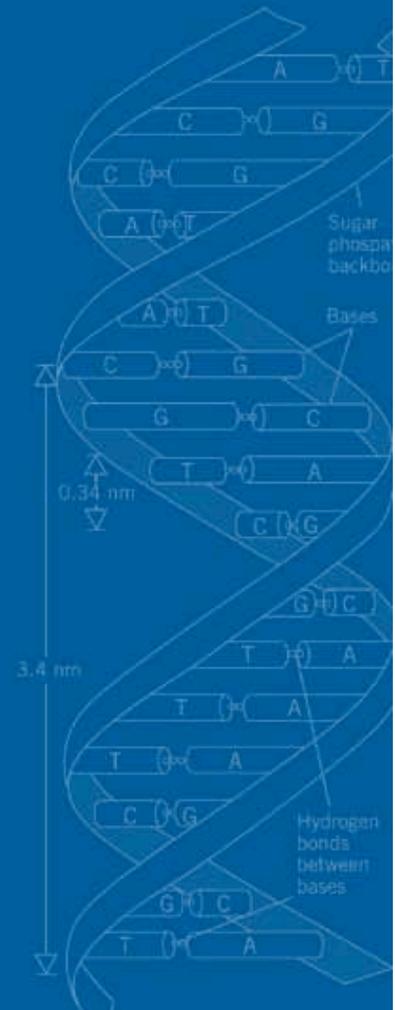
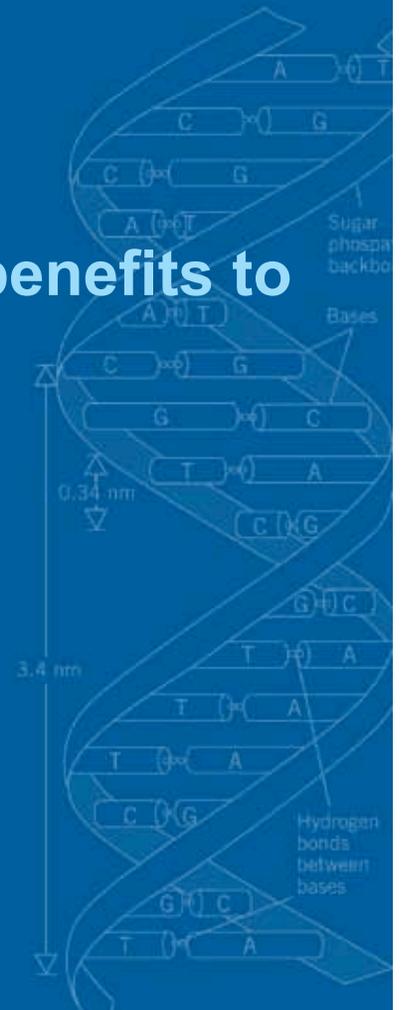


Incorporation of Fertility Endpoints in NHP Chronic Toxicology Studies for mAbs

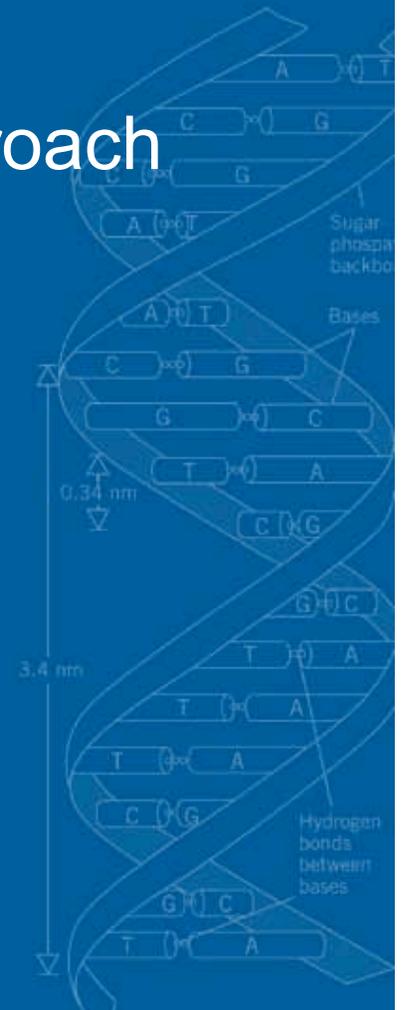
Anu Vaidyanathan, PhD, DABT
Toxicology/ Safety Assessment
Genentech, South San Francisco
May 6th, 2010



- Evolving change in industry
 - Many companies have paved the way
 - Helped us understand the challenges and benefits to using this approach
- Results in an optimized study design
- Follows ICH recommended 3 R's



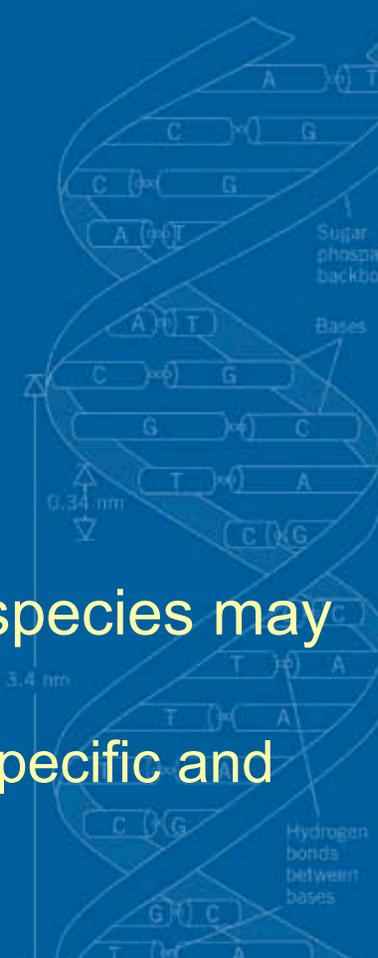
- ICH Regulatory Guidance
- Benefits & Strategy: A Combined Approach
- Alternative Approaches
- Conclusions



ICH S5 (R2), S6, M3 (R2)

- ICH S5 (R2): General guideline
 - At least one species, preferably rats
 - Fertility assessment
 - Pre-mating to conception
 - Mating behavior not assessed in NHP models
 - Conception to implantation
 - Not assessed in NHP models

❖ Issue: NMEs in which rat or alternative rodent species may not be appropriate (Korte et al 1987, 1993; Vogel & Bee 1999; Vogel 2000)
- Many monoclonal antibodies (mAbs) are highly specific and may not recognize rodent/rabbit/dog tissues



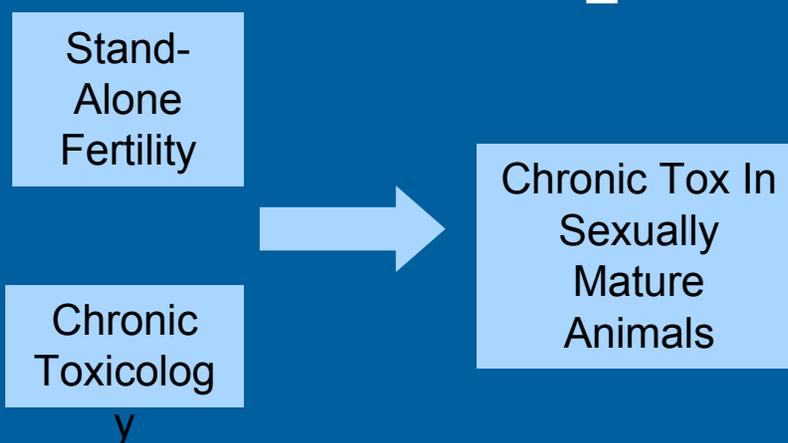
- ICH S6: Guidance for biotechnology-derived pharmaceuticals
 - **Study designs take into consideration:**
 - Species specificity, immunogenicity, half-life and biologic activity
 - Immunogenicity could impact PK/PD
 - **Determine target- dependent effects on fertility**
- Historical control database in both rodents and NHP
- ICH M3 (R2): Applies for recommended timing relative to clinical trials
 - **In general, completed prior to initiation of large scale or long duration trials (i.e. Phase III)**



❖ **Time, cost and resource investment prior to clinical POC**

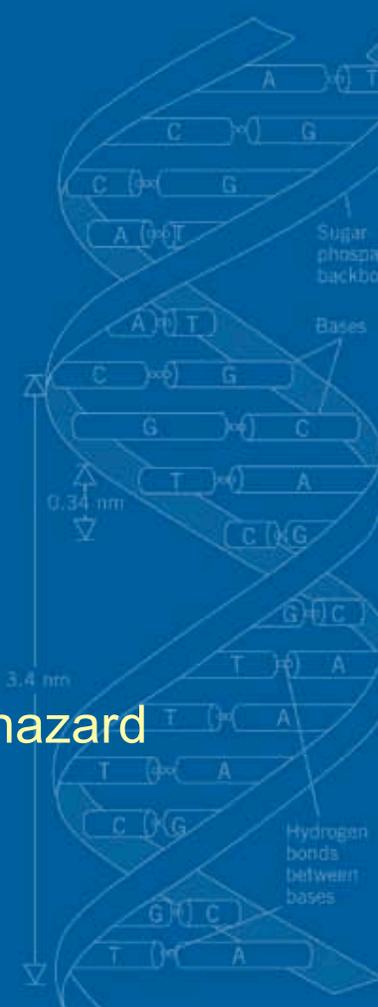
- ICH M3(R2) promotion of:

REDUCE **R**EFINE **R**EPLAC
E



A Combined Approach

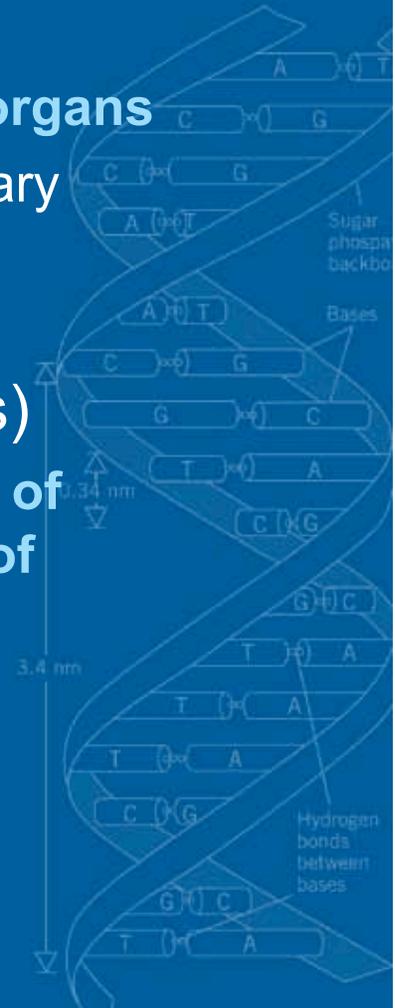
- Decreased animal usage = Less stress on animal supply
- Easily incorporate most endpoints used to identify fertility hazard
- Can generate multiple cycles of data
 - Menstrual or spermatogenic cycles



- Age and weight provide reasonable primary screen
 - **Males; ≥ 5 -6 ys; ≥ 5 kg (at study start)**
- Testicular volume (as measured by calipers/ultrasound)
- Functional parameters
 - **Serum testosterone, ejaculate volume, sperm parameters (count, motility, morphology)**
- Overall weight of evidence (WOE) approach using multiple factors
 - **Physical & functional parameters**

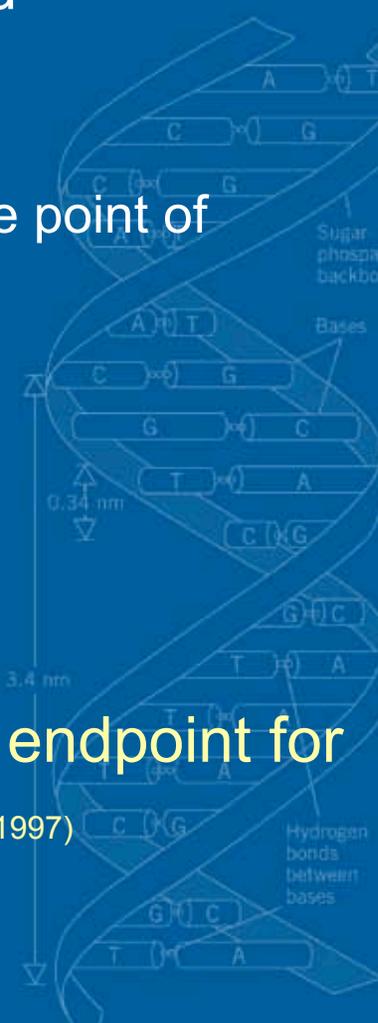
❖ **Ultimately collaborate with CROs experience & historical data to identify sexually mature animals**

- Easily incorporate most endpoints used to identify fertility hazard
 - ✓ **Organ wts & histo on reproductive and accessory organs**
 - Prostate, epididymis, seminal vesicles, adrenal & pituitary glands
 - Testes measurement
- Longitudinal effect (semen & hormone analysis)
 - ✓ **Morphology, sperm counts, sperm motility, volume of ejaculate (electro-ejaculation or at necropsy from tail of epididymis)**
 - Collection method could impact endpoints



- ✓ Spermatogenesis staging: Investigation of spermatid development
 - Represents a continuum of cellular differentiation
 - Identification of various germ cell types at a specific time point of development
 - Qualitative
 - Appropriately fixed and processed tissues
- Use weight of evidence (WOE)

❖ Histopathology is acknowledged as a sensitive endpoint for detecting fertility hazard (Sakai et al 2000; Takayama et al, 1995; Creasy, 1997)



- Age and weight provide reasonable primary screen
 - Females ≥ 4 yrs; 2.5 kg (post-pubertal at study start)
- Vaginal swabs
 - Duration of menstrual cycles? Normal vs Abnormal
 - Presence of regular menstrual cycles?
- Hormone analysis (E_2 /PG)
 - Look for evidence of ovulation
- Animals not enrolled based on cycle phase
 - Technically and logistically easier
- Overall WOE approach using multiple factors
 - Physical & functional parameters

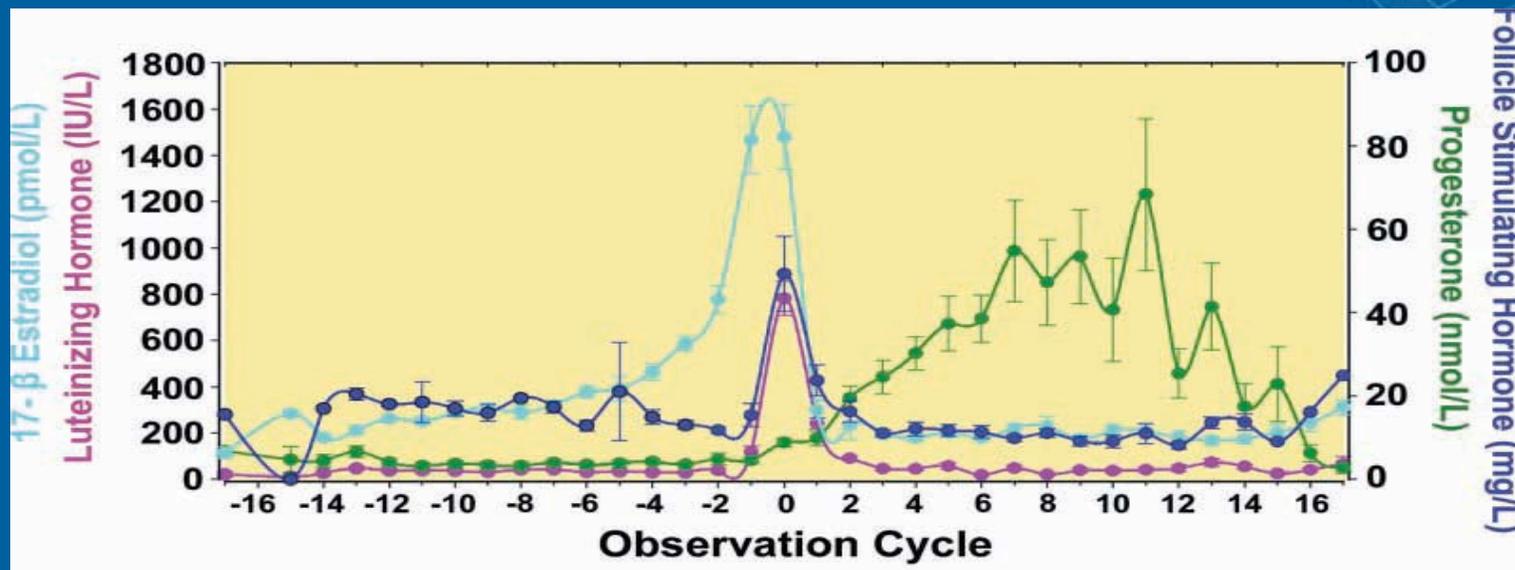
(Meyer et al, 2006; Smedley et al 2002; Weinbauer and Cooper 2000; Van Esch et al 2008)

❖ Ultimately collaborate with CROs experience & historical data to identify sexually mature animals

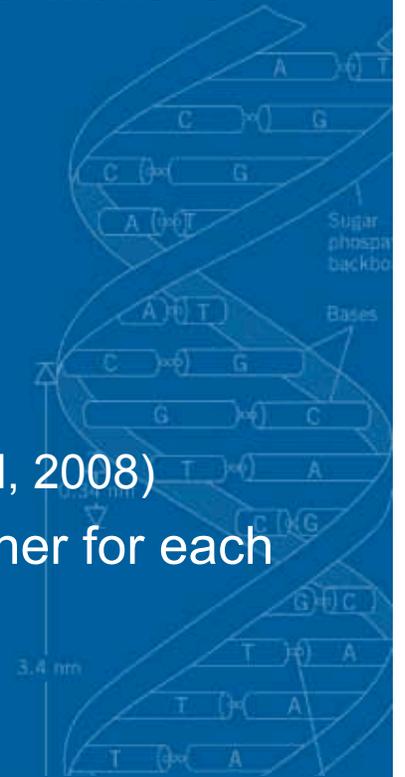


❖ Frequent blood sampling for female hormone cycling not appropriate for a chronic toxicology study

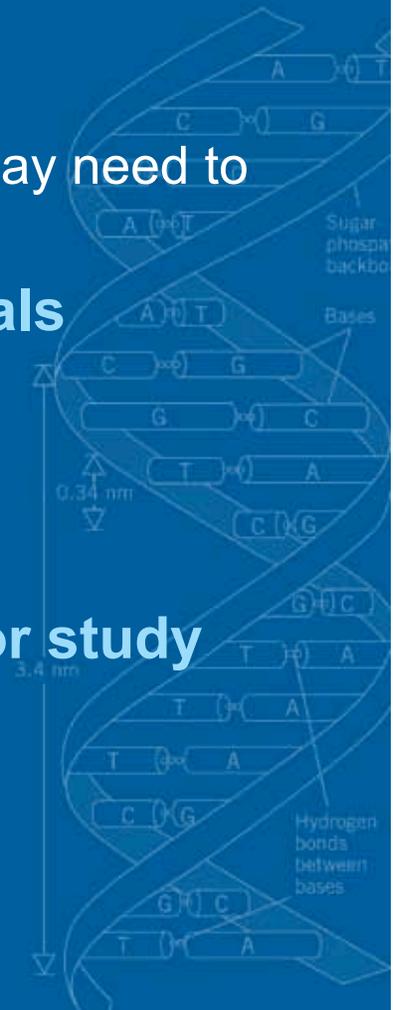
- For accurate/interpretable data need frequent blood sampling (~3 days/cyl)
 - Difficult to reconcile limited hormone data with other endpoints collected for fertility



- ✓ Daily vaginal swabs
 - Not evaluated for cytology, but for presence or absence of menses
 - Identifies cycle length
- Necropsy
 - ✓ Note general phase of cycle
 - ✓ Note sexual maturity status
 - Mammary glands
 - Presence or absence of multiple corpora luteum (CL)
 - Confirm appropriate endometrial morphology (Van Esch et al, 2008)
 - ✓ Microscopic evaluation of all reproductive tissues together for each animal
 - Integrated approach
- ❖ Most importantly, in the absence of hormone sampling, a WOE including vaginal swabs and histopathology can be used to detect fertility hazards



- **A combined approach**
 - **Overall development timeline impact?**
 - **Minimal**
 - Chronic study planning and implementation efforts may need to initiate sooner
 - **Depends upon supply of sexually mature animals**
 - **Increased screening efforts**
 - **Pre-study collections**
 - Physical and functional parameters
 - **Screen additional animals to meet required n for study**
 - **Additional cost**
 - **~\$170-200K**



Mating parameters generally not incorporated

- Low conception rates
 - 2 cycles of mating ~50-90%
 - Difficult to power adequately
- NHP have naturally low fertility rate and high spontaneous abortion rate
 - Abortion rates of 10-40%, many prior to GD 30
- Need ~90% reduction in sperm count to see effect on fertility
- No information on conception to implantation
- Not powered to detect infrequent events



Did we identify a hazard?

NO



- Proceed with development
- Additional nonclinical fertility studies not indicated

YES



- Combined approach geared toward hazard identification
 - Will not obtain hormone levels
 - Limited longitudinal data for individually affected animals

Follow-up Targeted Study (If not answered in combined approach)

- Characterize toxic insult
 - Is it consistent with MOA?
 - When did it occur?
 - Determine reversibility and monitorability
 - May need longer recovery phase to determine reversibility

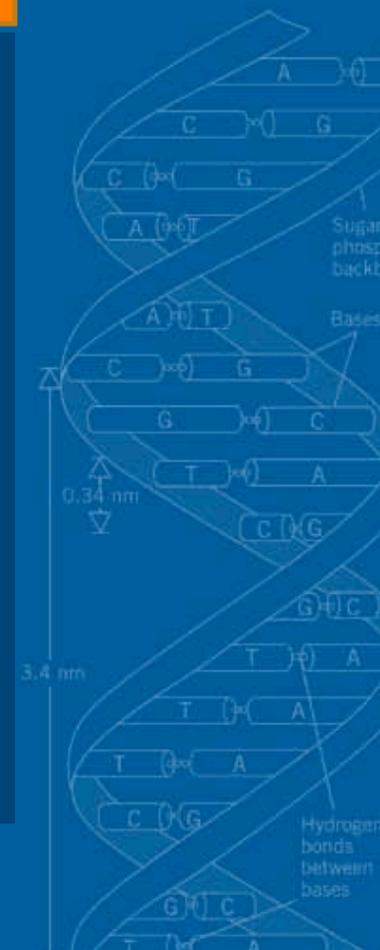
Patient

Population

- Key demographic
 - Non-life threatening disease
 - Reproductive years
- Oncology vs non-oncology
 - Early stage & adjuvant therapy?

MOA

- Mechanistic role in fertility?
 - Involvement in fertilization/implantation/placental development
 - Examples
 - Identify on target toxicity to understand mechanism

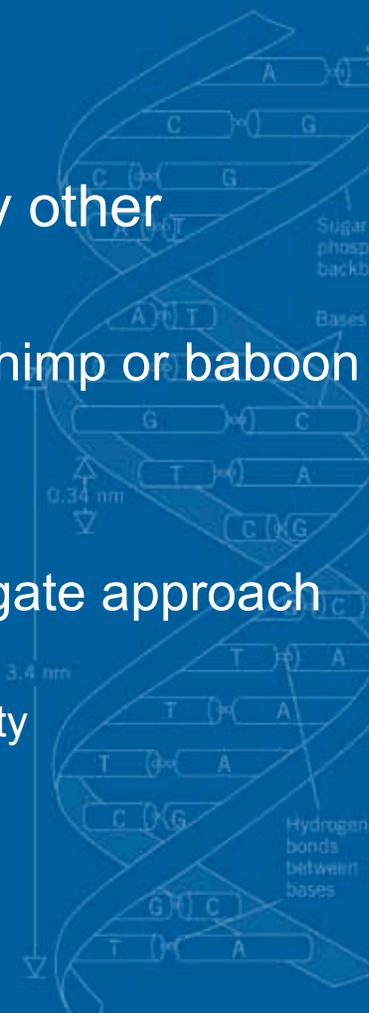


MOA= Mechanism of Action

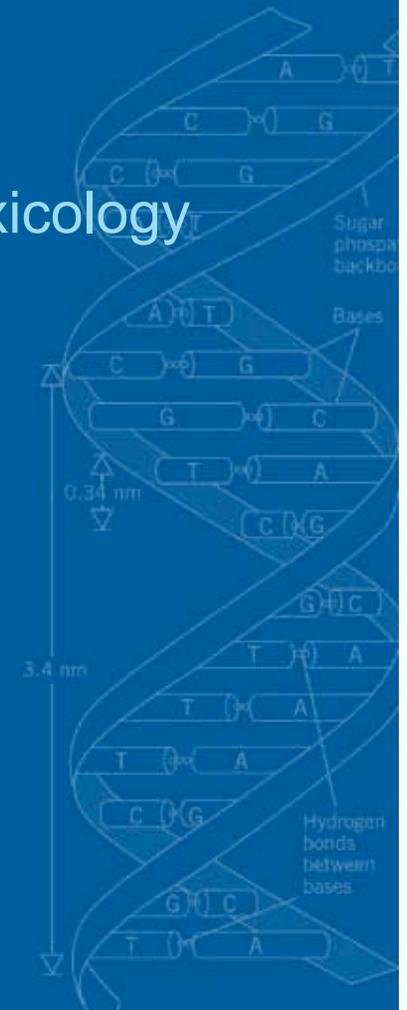
- Use of homologous Abs which cross reacts in rodent
 - **Pros**
 - Study design allows for obtaining data concerning:
 - Number of successful pregnancies
 - Impact on mating behavior
 - Conception and implantation
 - Multiple offspring
 - Shorter gestation, higher pregnancy rates and larger number of infants as compared to NHP
 - Larger number of animals per group to detect trends in the data



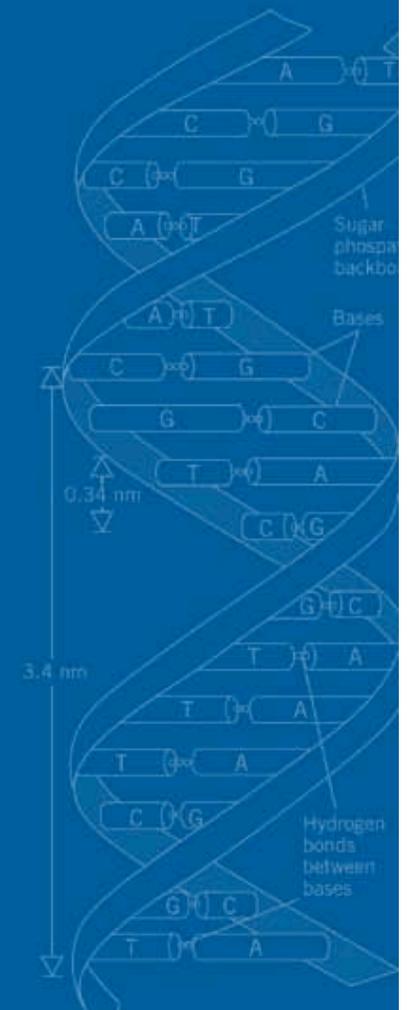
- Use of homologous Abs which cross reacts in rodent
 - **Cons**
 - Validate biology is representative
 - Relevant safety information can not be obtained by other methods
 - Less desirable unless only other binding species is chimp or baboon
 - Will not be testing your clinical candidate
 - Costly & time consuming (Martin et al 2009)
 - Additional validation efforts/assays required by surrogate approach
 - Biochemical characterization
 - Measurement of serum/plasma levels & potential immunogenicity
 - Separate manufacturing process



- Late stage development project
 - May have already conducted chronic tox study
- Considering the patient population & MOA
 - Were sexually mature animals used in general toxicology studies?
 - Histopathology on reproductive and accessory organs
- KO mating data available?
- Tissue cross reactivity
 - Binding in reproductive tissues?



- Clinical fertility endpoints may be acceptable by regulatory agencies
 - Good subject to address with agency
- Data generated
 - Endocrine, sperm and menstrual data
- Timing of fertility evaluation depends upon
 - Size of trial
 - Indication
 - Patient population
- May impact patient enrollment?



- Can easily incorporate most endpoints used to identify fertility hazard into chronic tox study designs
 - **May need follow-up if a fertility hazard is identified**
- Several criteria exist to ensure enrollment of sexually mature animals for general tox studies
 - **Ultimately rely upon CRO guidance**
- Alternative strategies exist, use case by case approach
 - **Consider stage of development**
 - **Patient population**
 - **MOA**
- Encourage open dialogue when possible with regulatory agencies when using alternative approaches



Key Contributors

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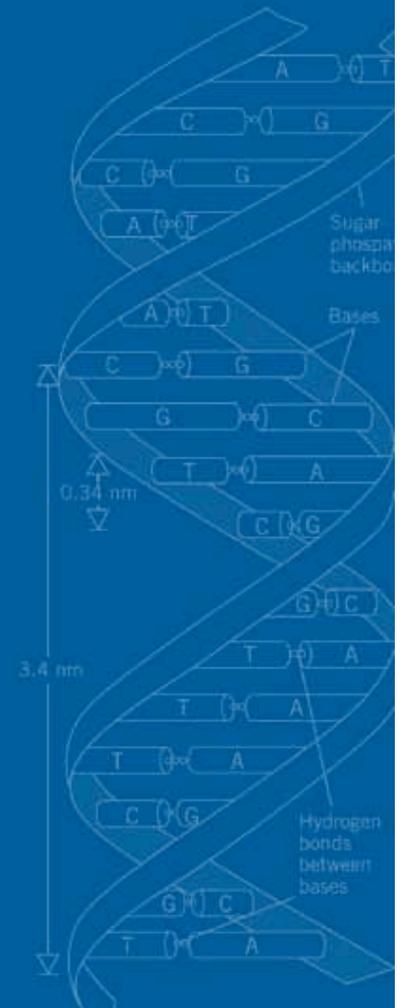
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Thought Leaders

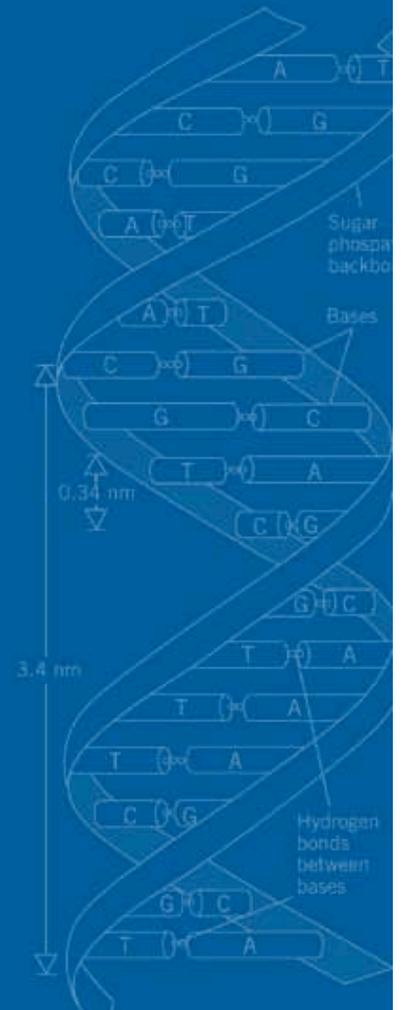
E Buse
R Korte
D Creasy
HC Dreef
J Vidal

S Rehm
F Vogel
TA Hendrie
AG Hendrickx
J Cavagnaro
A Hoberman

E Van Esch
GF Weinbauer
GJ Chellman
PL Martin
W Bee
JM Cline



Appendix



References

- Weinbauer GF, Niehoff M, Niehaus M, Srivastav S, Fuchs A, Van Esch E and Cline JM. Physiology and endocrinology of the ovarian cycle in Macaques. *Toxicol Pathol* 2008; 36; 7S.
- Van Esch E, Cline JM, Buse E, Weinbauer GF. The Macaque endometrium, with special reference to cynomolgus monkey (*Macaca fascicularis*). *Toxicol Pathol* 2008; 36; 67S.
- Van Esch E, Cline JM, Buse E, Wood CE, De Rijk EPCT, Weinbauer GF. Summary comparison of female reproductive system in human and the cynomolgus monkey (*Macaca fascicularis*). *Toxicol Pathol* 2008; 36:171S-172S.
- Weinbauer, GF. 2002. The non-human primate as a model in developmental and reproductive toxicity. In: Weinbauer, GF, Korte, R, editors. *Reproduction in nonhuman primates*. Munster, Germany: Waxmann Verlag GmbH. p 49-66.
- Weinbauer GF. 2005. Reproductive/developmental toxicity studies in the cynomolgus monkey: How close are we to human? In: Weinbauer GF, Buse E, Muller W, Vogel F, editors. *New developments and challenges in primate toxicology*. Munster, Germany: Waxmann Verlag GmbH. p 88-95.
- Weinbauer GF, Cooper TG. 2000. Assessment of male fertility impairment in the macaque model. In: Weinbauer GF, Korte R, editors. *Towards new horizons in primate toxicology*. Munster, Germany: Waxmann Verlag GmbH. p 13-42.
- Korte R, Weinbauer GF ed (1999). *Reproduction in nonhuman primates*. Munster, Germany: Waxmann Verlag GmbH.

References

- Vogel F. 2000. How to design male fertility investigations in the cynomolgus monkey. In: Weinbauer GF, Korte R, editors. *Towards new horizons in primate toxicology*. Munster, Germany: Waxmann Verlag GmbH. p 43-52.
- Vogel F, Bee W. 1999. Reproductive toxicology in primates: An overview of methods and techniques. In: Weinbauer GF, Korte, R, editors. *Reproduction in nonhuman primates*. Munster, Germany: Waxmann Verlag GmbH. P 95-110.
- Hendrie TA, Peterson PE, Short JJ, Tarantal AF, Rothgarn E, Hendrie MI, Hendrickx AG. Frequency of prenatal loss in a macaque breeding colony. *Am J Primatol* 1996; 40:41-53.
- Dreef HC, Van Esch E, De Rijk EPCT. Spermatogenesis in the cynomolgus monkey: A practical guide for routine morphological staging. *Toxicol Pathol* 2007; 35: 395-404.
- Meyer JK, Fitzsimmons D, Hastings TF, Chellman GJ. Methods for the prediction of breeding success in male cynomolgus monkeys (*Macaca fascicularis*) used for reproductive toxicology studies, *J Am Assoc Lab Animal Sci* 2006; 45:31-36.
- Smedley JV, Bailey SA, Perry RW, O'Rourke CM. Methods for predicting sexual maturity in male cynomolgus macaques on basis of age, body weight and histologic evaluation of the testes. *Contemp Top Lab Anim Sci* 2002; 41(5): 18-20.
- Creasy DM. Evaluation of testicular toxicology: A synopsis and discussion of the recommendations proposed by the Society of Toxicologic Pathology. *Birth Defects Research (Part B)* 2003; 68:408-415.
- Creasy, D. M. (1997). Evaluation of testicular toxicity in safety evaluation studies: The appropriate use of spermatogenic staging. *Toxicol Pathol* 25,119– 131.

References

- Sakai T, Takahashi M, Mitsumori K, Yasuhara K, Kawashima K, Mayahara H, Ohno Y. Collaborative work to evaluate toxicity on male reproductive organs by repeated dose studies in rats-Overview of the studies. *J Tox Sci* 2000; 25:1-21.
- Weinbauer GF, Frings W, Fuchs A, Niehaus M, Osterburg I. 2008. Reproductive/developmental toxicity assessment of biopharmaceuticals in nonhuman primates. In: Cavagnaro JA, editor. *Preclinical safety evaluation of biopharmaceuticals*. Hoboken, NJ: John Wiley & Sons. P 379-397.
- Takayama S, Akaike M, Kawashima K, Takahashi M, Kurokawa Y. A collaborative study in Japan on optimal treatment period and parameters for detection of male fertility disorders induced by drugs in rats. *J Amer College Tox* 1995; 14(4):266-292.
- Korte R, Vogel F, Osterburg I. The primate as a model for hazard assessment of teratogens in humans. *Arch Toxicol Suppl* 1987; 11: 115-121
- Korte R, Vogel F, Bee W, Osterburg I. 1993. The use of the non-human primate in reproduction toxicology. In: Korte R, Fanghanel J, Gossrau R, editors. *Teratologie- Embryologische Grundlagen, experimentelle und klinische Teratologie*. Berlin/ New York: Walter de Gruyter. P 167-191.
- Vogel F. 2000. How to design male fertility investigations in the cynomolgus monkey. In: Weinbauer GF, Korte R, editors. *Towards new horizons in primate toxicology*. Munster, Germany: Waxmann Verlag GmbH. p 43-51.
- Martin PL, Breslin W, Rocca M, Wright D, Cavagnaro J. Considerations in assessing the developmental and reproductive toxicity potential of biopharmaceuticals. *Birth Defects Res (Part B)* 2009; 86:176-203.

Utility of NHP Model for Fertility Assessment

Pros

- Physiology of menstrual cycle & endocrine systems of NHP similar to human
 - Similar parameters used to investigate fertility clinically
- Allows for collection of larger blood volumes
 - Potential use of biomarkers of toxicity for direct comparison to clinical endpoints

(Reviewed in Weinbauer 2005; Van Esch et al. 2008; Korte & Weinbauer 1999)

Cons

- Limited to assessment of estrous cycling and hormonal profiles
 - No data regarding mating behaviors, conception to implantation
- Generally underpowered as compared to rodent fertility studies

An example...

FEMALE STAND-ALONE NHP FERTILITY

3 mos cycle check

3 mos dosing phase

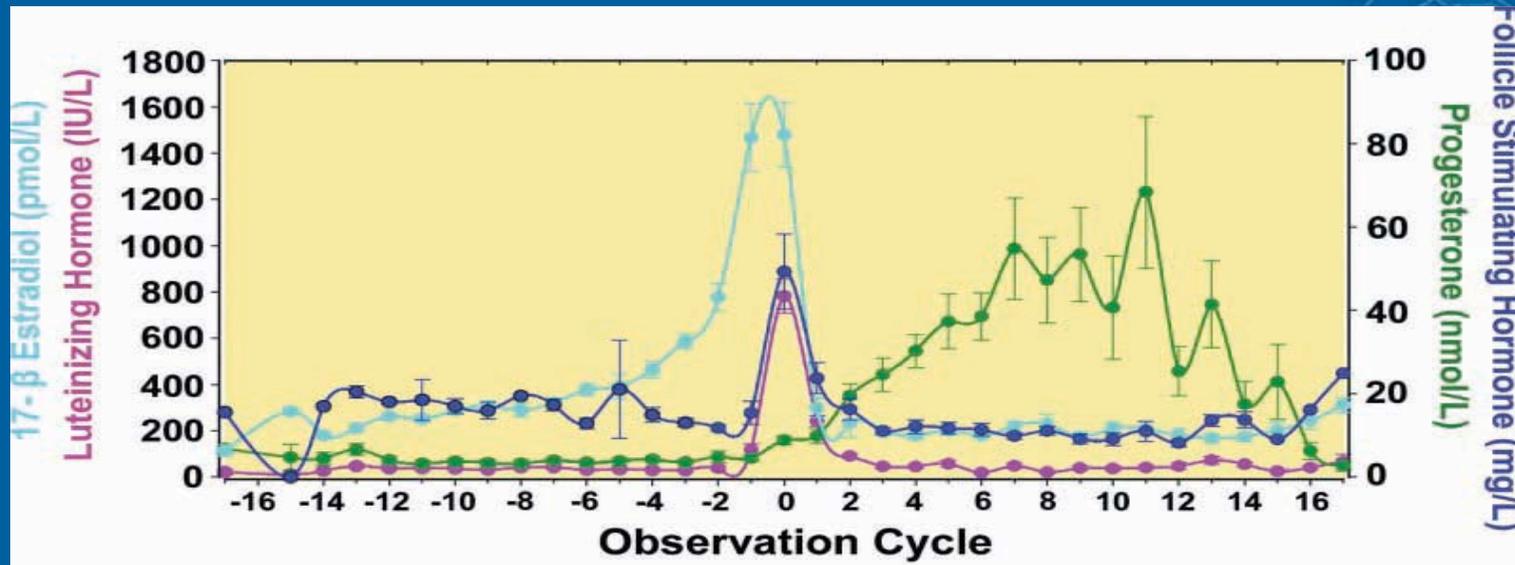
3 mos recovery*

Endpoints evaluated:

- ◆ Vaginal swabs
- ◆ Blood collections for estrogen and progesterone cycling data (every 3 days/cycle)

* Recovery dependant upon drug clearance
1 mens. cycle ~ 28 days +/- 8 days; consider synchronized dosing

- ◆ Histopathology- provides mechanism, histo may not add much value if no cycle abnormalities



Genentech
IN BUSINESS FOR LIFE

Follicular Phase

Ovulation

Luteal Phase

(Weinbauer et al., 2008)

An example...

MALE STAND-ALONE NHP FERTILITY

1 mo pre-dose ck

2 mos dosing phase

4 mos recovery*

Endpoints evaluated:

- ◆ organ wts & histo (prostate, epididymis, seminal vesicles, adrenal & pituitary glands)
- ◆ testes measurement
- ◆ semen analysis
- ◆ hormonal assessment (testosterone) (different labs have different capabilities)
- ◆ Spermatogenic staging

* Recovery dependant upon drug clearance

Justifications / Assumptions

- Dosing for 1 spermatogenic cycle ~40-45 days; epididymal transit time ~7-10 days (*Creasy, 2003*)

